

ETHNOPHARMACOLOGICAL STUDY OF SALT MARSH PLANTS FROM MUTHUKADU BACKWATERS

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INTRODUCTION

Salt marshes form in sheltered coastal areas where sediments accumulate and allow growth of angiosperm plants (Pennings & Bertness 2001) that comprise the foundation of the ecosystem. Salt marshes develop between terrestrial and marine environments, resulting in biologically diverse communities adapted for harsh environmental conditions including desiccation, flooding, and extreme temperature and salinity fluctuations. Marshes act as nurseries to a wide variety of organisms, some of which are notably threatened or marketed as important fisheries species.

Rapid growth of marsh vegetation and utilization of incoming nutrients make salt marshes highly productive systems, often yielding 2 kg of aboveground production per square meter, annually (Marinucci 1982, Dame 1989). In addition to providing habitat and food sources for many organisms, salt marshes benefit humans and surrounding ecosystems by sheltering coasts from erosion and filtering

nutrients and sediments from the water column.

Salinity in salt marshes is highly variable because of the influx of both fresh and saltwater into the environment. Freshwater enters upland marsh areas from terrestrial streams and rivers, increasing during periods of high precipitation. Saltwater inundates marshes during high tides, with dry seasons and high evaporation further increasing salinity. Salinity gradients caused by these processes contribute to zonation in marsh plants based on salt tolerance among species.

Most angiosperms have a limited ability to thrive in saline waters, and diversity of vegetation decreases with increasing salinity (Odum 1988, Odum & Hoover 1988). Seeds and seedlings are especially vulnerable to salt stress, further contributing to zonation in plants. However, many salt marsh plants have developed mechanisms to tolerate high salinities. Some plants increase succulence by retaining water or exclude salt at the roots, while others excrete salt through specialized

glands or sequester it into leaves that are shed periodically (Poljakoff- Mayer, 1975, Rozema et.al., 1981, Hacker and Bertness, 1995, Mitch and Gosselink, 1995, Dawes, 1998)

Since very old times, herbal medications have been used for relief of symptoms of disease [Maqsood, et.al., 2010]. Despite the great advances observed in modern medicine in recent decades, plants still make an important contribution to health care. Much interest, in medicinal plants however, emanates from their long use in folk medicines as well as their prophylactic properties, especially in developing countries. Large number of medicinal plants has been investigated for their antioxidant properties. Natural antioxidants either in the form of raw extracts or their chemical constituents are very effective to prevent the destructive processes caused by oxidative stress [Zengin, 2011]. Although the toxicity profile of most medicinal plants have not been thoroughly evaluated, it is generally accepted that medicines derived from plant products are safer than their synthetic counterparts [Vongtau, *et.al.*, 2005 and Oluyemi, *et.al.*, 2005].

A large proportion of the human population depends on traditional medicine. Medicinal plants have become the focus of intense study recently in terms of conservation and as to whether their traditional uses are supported by actual pharmacological effects or merely based on folklore. With the increasing acceptance of traditional medicine as an alternative form of health care, the screening or medicinal plant for active compound is very important. The situation is alarming in developing as well as developed countries due to indiscriminate use of antibiotics. The drugresistant bacteria and fungal pathogens have further complicated the treatment of

infectious diseases in immune compromised, AIDS and cancer patients [Diamond, 1993]. It is likely that plant extract showing target sites other than those used by antibiotics will be active against drug-resistant microbial pathogen. However, very little information is available on such activity of medicinal plants (Lee, *et al.*, 1998).

Plants, as the source of medicine, have been playing an important role in the health services around the globe [Thomson, 2010]. About three quarters of the world's population relies on plant and their extracts for health care [Kunwar and Bussmann, 2008]. A good number of our population particularly those living in rural areas depend largely on herbal remedies for the treatment of different types of diseases. It indicates the importance of the individual plants in the health care system.

Ethnopharmacology can be defined as the interdisciplinary scientific exploration of biologically active agents traditionally employed or observed by man [Holmstedt, 1995]. Its objectives are to rescue and document an important cultural heritage as well as evaluate the agents employed [Holmstedt, and Bruhn, 1995]. One common approach in this field is a literature search using several published genetic resources. The application of new bioinformatics database systems about herbal texts holds great promise for identifying novel bioactive compounds for pharmacotherapy [Buenz, *et.al.*, 2004]. Some International Databases, such as Natural Products Alert (<http://www.napralert.org/Default.aspx>), provide information about pharmacological activities, ethnopharmacological data, chemical compounds and data from tests on animals and humans for thousand of species from all over the world. Phyto chemistry can contribute to the synthesis of new drugs with

therapeutic properties [Naranzo, 1995]. Nature provides enormous potential for the discovery of new bioactive compounds; at least a million different compounds could be isolated [Verpoorte, 1998].

For a long period of time in history, plants have been valuable and indispensable sources of natural products for the health of human beings and they have a great potential for producing new drugs (Nascimento, 2008; Littleton, 2005). Even today people who live near to the forests use plant products to cure chronic diseases. Tropical and subtropical areas of the world are bestowed with abundant flora and herbs which have untapped properties, such as antimicrobial, antiviral and antifungal. According to the World Health Organization, plants are a source of compounds that have the ability to combat disease, antimicrobial, antiviral and antifungal activities (Gazim, 2008). In addition, medicinal plants have been used for centuries as remedies for human ailments and diseases because they contain components of therapeutic value (Panda, 2009). Also they are less toxic to humans

AVICENNIA MARINA



Scientific Name: *Avicennia marina* (Forrsk.)Vierh

Synonym: *Sceura marina* Forssk

Local Name: Qurm, Gurm

Arabic Name(s): Shorah, Qurm, Mangrove

Common name: Mangrove, Grey mangrove, Tivar

Family: Avicenniaceae (Verbenaceae)

Description of the Plant:

and environmentally friendly due to fewer pollutants produced in production and have minimal health hazards (Opra and Wokocho, 2008). However literature related to the ethno-medicinal importance of salt marsh plants are scarce, knowledge of the chemical constituents of plants is desirable, not only for the discovery of therapeutic agents, but also because such information may be of value in disclosing new sources of such economic materials as tannins, oils, gums, precursors for the synthesis of complex chemical substances, etc. In addition, the knowledge of the chemical constituents of plants would further be valuable in discovering the actual value of folkloric remedies (Farnsworth, 1966).

The aim of the study is to elaborate the ethnopharmacology of salt marsh plants in this case, 6 variety of plants (*Avicennia marina*, *Avicennia officinalis*, *Sesuvium prostracastrulam*, *Salicornia branchiate*, *Suaeda maritime*, and *Suaeda monoica*) found in the shore of Muthukadu back waters, Tamil Nadu.

Small evergreen tree, up to 10m high, stem erect with fine pale gray scales. Leaves simple leathery, opposite, ovate, petiolate with entire margin and acute tip, dark glossy green on the upper surface, dull greyish on the lower surface with excreted salt crystals. As *Avicennia* is growing in a specialized habitat, which is poorly aerated, it is adapted to life in this habitat by the presence of erect leafless outgrowths of the roots called pneumatophores or breathing

AVICINNEIA OFFICINALIS



Family- Avicenniaceae

Habitat- This species is found in Bangladesh, India, Indonesia, Malaysia, Brunei, Myanmar, Philippines, Singapore, Sri Lanka, Thailand, Viet Nam, and southern Papua New Guinea

English- Grey Mangrove, White Mangrove

Folk: Kanna (Tamil), Tavarian (Gujarat), Orayi (Malayalam)

Description of the plants:

The flower, the largest among the *Avicennia* species has a diameter of 6 to 10 mm when expanded. It is orange yellow to lemon yellow in color.

Action- Fruits are plastered on to boils and tumours, poultice of unripe seed stop inflammation, roots used for its aphrodisiac, bark is used to treat skin problems especially scabies, resin for snake bite and

roots up to 50 cm long, they stick out above water and absorb air, which thought to oxygenate the roots.

Action:

Bark astringent and used as aphrodisiac, for scabies, antifertility agent and has tanning properties. Flowers for perfumes. Leaves are aphrodisiac and used for toothache, Leaves and seeds forage for camels and animals.

contraceptive by women, seed for ulcers. This plant contains pentacyclic triterpenoids such as lupeol, betulin, betulinaldehyde, betulinicacid, beta-sitosterol and Iridoid glucosides having c-11 carboxylic acidgroup were also present and other compounds present are flavanoids ,alkaloid, steroids, tannins, wax esters are the most considerable compounds.

SESUVIUM PORTULACASTRUM



Family: Aizoaceae

Habitat: Coastal dunes and beaches, Worldwide in Tropical and Subtropical regions.

English: Sea Purslane, Shoreline Purslane

Folk: Dhapa, Orputu, Vancaravacci, Vangaredukura, Jadu palang

Description of Plant: *Sesuvium portulacastrum* is a sprawling perennial herb up to 30 centimetres (12 in) high, with thick, smooth stems up to 1 metre (3.3 ft) long. It has smooth, fleshy, glossy green leaves that are linear or lanceolate, from 10–70 millimetres (0.39–2.76 in) long and 2–15 millimetres (0.079–0.591 in) wide. Flowers are pink or purple.

Action:

It has been utilized for the treatment of epilepsy, conjunctivitis, dermatitis, haematuria, leprosy and purgative, toothache and also as antimicrobial agent. Extract of this plant and the essential oil from the fresh leaves of *S. portulacastrum* showed antibacterial, antifungal as well as antioxidant activity. The ethanolic extract of the medicinal plant *S. portulacastrum*

showed potential against the causative agents and pathogens related to various gastrointestinal disorders leading to indigestion, dysentery, and diarrhoea. Moreover the ethanolic extract of the medicinal plant *S. portulacastrum* showed potential against the causative agents of nosocomial infections, *Staphylococcus aureus* and *E. coli*.

SUAEDA MARITIMA



Family: Amaranthaceae

Folk: Sawad, Umiri in Pitchvaram, Mattaumiri in Muthupet, Tamil nadu

English: Common Sea Blite, Shrubby Sea Blite

Habitat: It is the shrub with continuous unjoined stems found in western region of Saudi Arabia [Yousef, *et.al.*, 2009]. Both in the Pichavaram and the Muthupet mangroves *Suaeda maritima* can be seen growing as monospecific patches in large areas in coup felled areas.

Action:

The juice of this herb is used for treatment of liver diseases by Arab practitioners [Patra, *et.al.*, 2011]. The leaves are also used as remedy for liver, heart, and lipid disorders [Lin, *et.al.*, 2008]. The ethanolic extracts of *S. maritima* leaves significantly

attenuated concanavalin (a hepatotoxin) induced biochemical (serum AST, ALT, APT, and bilirubin) and histopathological changes in liver [Ravikumar, *et.al.*, 2011]. The extract of plant also showed significant antioxidant, anti-inflammatory, antiviral,

and antibacterial activities [Ravikumar, *et.al.*, 2011 and Singh, *et.al.*, 2013] which may contribute to its hepatoprotective activity. It is nontoxic edible plant which is used in salad and as fodder for animals. The LD₅₀ of ethanolic extract of *S. maritima* in

rats was found to be 3 g/kg b.w. [Ravikumar, *et.al.*, 2011]. Phytochemical studies on plant of *S. maritima* showed the presence of alkaloid, flavonoid, sterols, phenolic compounds, and tannins [Singh, *et.al.*, 2013] .

SUEADA MONOICA



Family: (Chenopodiaceae)

Habitat: is a salt marsh mangrove herb similar to *Suaeda maritima* (L). Dumort in appearance, growing in hypersaline soils. It is distributed throughout the East west coast mangroves in India viz., Sunderbans in West Bengal state, Bitharkanika and Mahnadhi in Orissa state, Coringa, Godavari and Krishna in Andhra Pradesh State, Pichavaram, Muthukadu, Karangadu and Muthupet in Tamil Nadu state.

Folk: Vellaikirai (or) Nilavumari (seaside Indian salt wort).

Description of Plant: It is a shrub but much smaller in size (0.3-0.7mm in length) when compared to *Suaeda maritima*. Leaves simple, succulent, linear, young twigs are slender ribbed.

Action:

The leaves have been used as edible green leaves. The ash obtained from burnt plant parts have been exported without knowing the purpose. Traditionally, the leaf from *Suaeda monoica* is known to use as a medicine for hepatitis [Bandarnayake, 1998] and scientifically it is reported to be used as ointment for wounds [Padmakumar and Ayyakannu, 1992] and possess antiviral

activity [Premanathan, *et.al.*, 1992] because of the presence of triterpenoids, sterols [Ghosh, *et.al.*, 1985 and Subramanya, *et.al.*, 1992]. The present attempt has been made to find out the hepatoprotective evaluation of crude ethanolic extract from leaves of *S. monoica* for possible development of hepatoprotective herbal medicine.

SALICORNIA BRACHIATA



Family: Chenopodiaceae

Habitat: Throughout India, Sunderbans, Pichavaram and Mudhupet and in Sri Lanka - Jaffna, Hambantota and Kirinda.

Folk: Kozhikal in Pichavaram, Pavazhappoondu in Muthupet, Batura, Katula, Kattu Umari

Description of plant:

Herb, head erect or decumbent of about 20 to 45 cm high. Stems succulent and much branched. Each segment of the stem from a little cup at the apex. The "cup" has short teeth covering the base of the next segment. Fruits utricle, ovoid, membranous, enclosed in spongy perianth; seed pale brown, hispid with white hairs.

Action:

Ash of this plant is used to cure itches and the leaf and stem extracts are used for treating hepatitis. The extracts of leaf of *S. brachiata* showed potent antioxidant

activity. The plants rich in tannins have significant activity in cancer prevention and are used in treating intestinal disorders. Flavonoids are known to possess a wide range of biological activities such as antioxidant, antimicrobial, anti-inflammatory and anticancer activities. Several species of *Salicornia* possess antibacterial and antihypertensive properties and are quoted in folk medicine for relief of toothache and chronic rheumatism (Rizk, 1986), constipation, obesity, diabetes and cancer.(Park, 2000 and Deepa *et.al.*, 2013)

BIOACTIVE COMPOUNDS FROM MANGROVES

The common chemical constituents present in the mangroves are aliphatic alcohols and acids, amino acids, alkaloids, carbohydrates, carotenoids, hydrocarbons, free fatty acids including polyunsaturated fatty acids, lipids, pheromones, phorbol esters, phenolics and related compounds, steroids, triterpenes and their glycosides, tannins and other terpenes [Revathy, *et.al.*,

2013]. Even though several chemical studies have been conducted on mangrove plants, reports pertaining to their activity-structure relationship are very few. Some common salty marsh plants found in tropical and sub tropical regions, their traditional uses, general chemical constituents, *in vitro* bioactivity etc are given in Table 1.

Table 1: Common mangroves with *in vitro* bioactivity

S.No	Mangroves	Traditional uses	General Chemical composition	<i>In vitro</i> activity	Reference
1.	<i>Avicennia marina</i>	cure for skin diseases	terpenoids, steroids naphthalene derivatives, flavones, glucosides	cytotoxic antibacterial antifungal antioxidant	Fauvel <i>et.al.</i> , 1993 Gurudeeban, <i>et.al.</i> , 2012 Han L, Huang XS, 2008 Feng Y <i>et.al.</i> , 2006
2.	<i>Avicennia</i>	aphrodisiac, diuretic,	arsenic,	antibacterial	Bandaranayake,

	<i>officinalis</i>	cure for hepatitis, leprosy,	alkaloids, saponins, tannins, tri terpenoids	anti-ulcer	2002 Sharma and Garg, 1996
3.	<i>Suaeda maritima</i>	Cure for hepatitis Immunity booster	carbohydrate, alkaloids, glycosides, flavonoids sterols, phenolic and tannins compounds.	antiviral, antibacterial activity, anti-inflammatory and antioxidant	Patra et.al., 2011 Singh et.al., 2013 Bandaranayake, 1998 Ravikumar, et.al., 2011
4.	<i>Sesuvium portulacastrum</i>	remedy for fever, kidney disorders ,scurvy and in the treatment of various infections,hepatoprotective activities, epilepsy, conjunctivitis, dermatitis, haematuria, leprosy and purgative, toothache	saponins, alkaloids, polyphenols, terpenoids	antibacterial and anticandidal activities and moderate antifungal activity. Antioxidant, to treat various gastrointestinal disorders leading to indigestion, dysentery, and diarrhoea, fights against nosocomial infections	Kokpal, et.al., 1990 Robert and Frank, 1997 Michael, et.al., 2006 Chandrasekaran et.al., 2011
5.	<i>Suaeda monoica</i>	medicine for hepatitis, ointment for wounds and possess antiviral activity	triterpenoids and sterols, saponins, phenols, alkaloids	antiviral activity, antibacterial, anti-inflammatory, anti-oxidant	Bndarnayake, 1998 Padmakumar and Ayyakannu, 1992 Premanathan, et.al., 1992 Ravikumar, et.al., 2010
6.	<i>Salicornia branchiata</i>	to treat itches	High amount of phenols and flavonoids,	Antibacterial, antiallergic, anti-inflammatory, antimicrobial,	Kathiresan, et.al., 2001 Ravindran,

				antiviral, antioxidant, oestrogenic, enzyme inhibition, vascular and cytotoxic antitumour activity	et.al., 2005 Manikandan, et.al., 2005 Stanley, 2008
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DISCUSSION

From this review study, it is clear that the medicinal plants play a significant role against on various diseases. Different medicinal herbs and plants extracts have potent hepatoprotective activity in various animal models. The hepatoprotective activity is probably due to the presence of flavonoids, phenolic compounds, polyphenols etc in all few herbal plants. The results of this study indicate that extracts of leaves and plants extracts of some medicinal plant have good potentials for use in hepatic disease.

CONCLUSION

A phytotherapeutic approach to modern drug development can provide many invaluable drugs from traditional medicinal plants. Search for pure phytochemicals as

drugs is time consuming and expensive. Numerous plants and polyherbal formulations are used for the treatment of liver diseases. However, in most of the severe cases, the treatments are not satisfactory. Although experimental evaluations were carried out on a good number of these plants and formulations, the studies were mostly incomplete and insufficient. The therapeutic values were tested against a few chemicals-induced subclinical levels of liver damages in rodents. Development of such medicines with standards of safety and efficacy can revitalize treatment of liver disorders and hepatoprotective activity.

REFERENCES

1. Bauer A.W, Kirby W.M.M, Sherris J.C, and Turck M, Antibiotic Susceptibility Testing by a Standardized Single Disk Method, American Journal of Clinical Pathology, 45, 1996, 493–496.
2. Cushnie TP, and Lamb AJ, Antimicrobial activity of flavonoids, Int J Antimicrob Agents, 26 (5), 2005, 343-56.
3. Farnsworth N.R, Biological and phytochemical screening of plants, J. Pharm. Sci., 55, 1966, 225-276.
4. Gazim, Rezende Z.C, Fraga C.M, Svidzinski S.R, Cortez T.I, Antibacterial activity of the essential oil from *Calendula officinalis* L.(Asteracea) growing in Brazil, Braz J Microbiol., 39, 2008, 61–3.
5. Gibbs R.D, Chemotaxonomy of flowering plants, McGill queen's university press, Montreal, 1974, 523-619.

6. Harborne J.B, Phytochemical methods, London, Chapman and Hall Ltd., 1973, 49-188. Littleton, Rogers J, Falcone T, Novel approaches to plant drug discovery based on high throughput pharmacological screening and genetic manipulation, *Life Sci.*, 78, 2005, 467–75.
7. Nascimento G.G.F, Locatelli, Freitas J, Silva P.C, Antibacterial activity of plant extracts and phytochemicals on antibiotic resistant bacteria, *Braz J Microbiol.*, 31, 2000, 247–56.
8. Oakenfull, Fenwick, Saponin content of soybeans and some commercial soybean products, *J.Sc Food Agric.*, 32, 1981, 273-278.
9. Omojate Godstime, Enwa Felix O, Jewo Augustina O, Eze Christopher O, Mechanisms of Antimicrobial Actions of Phytochemicals against Enteric Pathogens – A Review, *J Pharm Chem Biol Sci.*, 2 (2), 2014, 77-85.
10. Opra E.U, Wokocha R.C, Efficacy of some plant extracts on the in vitro and in vivo control of *Xanthomonas campestris* P.v. *Vesicatoria*, *Agric J.*, 3, 2008, 163–70.
11. Panda S.K, Thatoi H.N, Dutta S.K, Antibacterial activity and phytochemical screening of leaf and bark extracts of *Vitex negundo* I. from Similipal biosphere reserve, Orissa, *J Med Plant Res.*, 3, 2009, 294–300.
12. Holmstedt, B. In: *Ethnobotany: Evolution of a Discipline*; Schultes, R.E.; von Reis, S.; Eds.; Timber Press Inc.: Oregon, **1995**, pp. 320-337.
13. Holmstedt, B.; Bruhn, J.G. In: *Ethnobotany: Evolution of a Discipline*; Schultes, R.E.; von Reis, S., Eds.; Timber Press Inc.: Oregon, **1995**, pp. 338-342.
14. Buenz, E.J.; Schnepple, D.J.; Bauer, B.A.; Elkin, P.L.; Riddle, J.M.; Motley, T.J. Techniques: Bioprospecting historical herbal texts by hunting for new leads in old tomes. *Trends Pharmacol.Sci.*, **2004**, 25, 494-498.
15. Naranjo, P. In: *Ethnobotany: Evolution of a Discipline*; Schultes, R.E.; von Reis, S., Eds.; Timber Press Inc.: Oregon, **1995**, pp. 362-368.
16. Verpoorte, R. Exploration of nature's chemodiversity: the role of secondary metabolites as leads in drug development. *Drug Discov.Today*, **1998**, 3, 232-238
17. Diamond RD., 1993. The growing problem of mycoses in patients infested with human immunodeficiency virus. *Review of Infectious Diseases*, 13: 480-486.
18. Lee CK., Kin H., Moon KH., Shun KH., 1998. Screening and isolation of antibiotic resistance inhibitors from herb material resistance inhibition of volatile components of Korean aromatic herbs. *Archives of Pharmaceutical Research*, 2: 62-66.
19. Maqsood S, Singh P, Samoon MH, Balange AK: Effect of dietary chitosan on non-specific immune response and growth of *Cyprinus carpio* challenged with *Aeromonas hydrophila*. *Inter Aqua Res* 2010, 2:77–85.
20. Zengin G, Cakmak YS, Guler GO, Aktumsek A: Antioxidant properties of methanolic extract and fatty acid composition of *Centaurea urvillei* DC. subsp. *hayekiana* Wagenitz. *Rec Nat Prod* 2011, 5:123–132.
21. Vongtau HO, Abbah J, Chindo BA, Mosugu O, Salawu AO, Kwanashie HO, Gamaniel KS: Central inhibitory effects of the methanol extract of *Neorautanenia mitis* root in rats and mice. *J Pharm Biol* 2005, 43:113–120.
22. Oluyemi KA, Okwuonu UC, Baxter DG, Oyesola TO: Toxic effects of methanolic extract of *Aspilia africana* leaf on the estrous cycle and uterine tissues of Wistar rats. *Int J Morphol* 2007, 25:609–614.
23. Thomson GE, Further consideration of Asian Medicinal plants in treating common chronic disease in West. *Journal of Medicinal Plants Research*, 4(2) (2010) 125.

24. Kunwar RM & Bussmann RW, Ethnobotany in the Nepal Himalaya. *Journal of Ethnobiology and Ethnomedicine*, 4(24) (2008).
25. Adam, P. 1990. *Saltmarsh ecology*. Cambridge University Press. Cambridge, UK. 461 pp.
26. Anderson, CE. 1974. A review of structures of several North Carolina salt marsh plants. In: Reimold, RJ & WH Queen, eds. *Ecology of halophytes*. 307-344. Academic Press. New York. USA.
27. Armstrong, W. 1979. Aeration in higher plants. *Adv. Bot. Res.* 7: 225-332.
28. Brockmeyer JR., RE, Rey, JR, Virnstein, RW, Gilmore, RG & L Earnest. 1997. Rehabilitation of impounded estuarine wetlands by hydrologic reconnection to the Indian River Lagoon, Florida (USA). *Wetlands Ecol. Manag.* 4: 93-109.
29. Chapman, VJ. 1960. *Salt marshes and salt deserts of the world*. Leonard Hill Limited. London, UK.
30. Coles, SM. 1979. Benthic microalgal populations on intertidal sediments and their role as precursors to salt marsh development. In: Jefferies, RL & AJ Davy, eds. *Ecological processes in coastal environments*. 25-42. Blackwell Scientific Publications. Oxford, UK.
31. Costa, CSB & AJ Davy. 1992. Coastal salt marsh communities of Latin America. In: Seeliger, U, ed. *Evolutionary ecology in tropical and temperate regions: coastal plant communities of Latin America*. 179-199. Academic Press. San Diego, CA. USA.
32. Crewz, DW & RR Lewis III. 1991. *An evaluation of historical attempts to establish vegetation in marine wetlands in Florida*. Florida Sea Grant technical paper TP-60. Sea Grant College, University of Florida. Gainesville, FL. USA.
33. Dame, RF. 1989. The importance of *Spartina alterniflora* to Atlantic coast estuaries. *Rev. Aquat. Sci.* 1: 639-660.
34. David, JR. 1992. *The Saint Lucie County Mosquito Control District summary workplan for mosquito impoundment restoration for the salt marshes of Saint Lucie County*. Saint Lucie County Mosquito Control District. Saint Lucie, FL. USA.
35. Dawes, CJ. 1998. *Marine botany, 2nd ed.* John Wiley & Sons. New York. USA. 480 pp.
36. Drake, BG. 1989. Photosynthesis of salt marsh species. *Aquat. Bot.* 34: 167-180.
37. Dybas, CL. 2002. Florida's Indian River Lagoon: an estuary in transition. *BioScience*. 52: 554-559.
38. Eleuterius, LN & CK Eleuterius. 1979. Tide levels and salt marsh zonation. *Bull. Mar. Sci.* 29: 394-400.
39. Field, DW, Reyer, AJ, Genovese, PV & BD Shearer. 1991. *Coastal wetlands of the United States*. National Oceanic and Atmospheric Administration and US Fish and Wildlife Service. Washington, DC.
40. FNAI. 1997. *County distribution and habitats of rare and endangered species in Florida*. Florida Natural Areas Inventory. Tallahassee, FL. USA.
41. Flowers, TJ, Troke, PF & AR Yeo. 1977. The mechanism of salt tolerance in halophytes. *Ann. Rev. Plant Physiol.* 28: 89-121.
42. Flowers, TJ, Hajibagheri, MA & NWJ Clipson. 1986. Halophytes. *Q. Rev. Biol.* 61: 313-337.
43. Ford, MA & JB Grace. 1998. Effects of vertebrate herbivores on soil processes, plant biomass, litter accumulation and soil elevation changes in a coastal marsh. *J. Ecol.* 86: 974-982.

44. FWS. 1999. Coastal Salt Marsh. *In: Multi-species recovery plan for South Florida*. US Fish & Wildlife Service. 553-595.
45. Hacker, SD & MD Bertness. 1995. A herbivore paradox: why salt marsh aphids live on poor-quality plants. *Amer. Nat.* 145: 192-210.
46. Lakshmanan*, C. Rajeshkannan, A. Kavitha, B. Mekala and N. Kamaladevi, Preliminary screening of biologically active constituents of *Suaeda monoica* and *Sesuvium portulacastrum* from palayakayal mangrove forest of Tamilnadu G. JPP 2 (3), 2013, 149-152
47. Sathish P, Jaswanth G, Gurudhathan K.B, Gopinath J, Gayathri P.K, Yuvaraj D, Phytochemical investigation and antibacterial activity of salt marsh plant extracts, *Journal of Chemical and Pharmaceutical Sciences*, ISSN: 0974-2115, JCHPS 9.,(1) 67, March 2016, 292-294.
48. Robert IL, Frank W. The biological flora of Coastal and wetlands *Sesuvium portulacastrum* LJ. *Coast Res* 1997; 13:96-104.
49. Lonard RI, Judd FW, Coast LJ. The biological flora of coastal dunes and wetlands. *Sesuvium portulacastrum*, *Res* 1997; 13(1):96-104.
50. Chandrasekaran M, Senthilkumar A, Venkatesalu V, Antibacterial and antifungal efficacy of fatty acid methyl esters from the leaves of *Sesuvium portulacastrum* L. *European Review for Medical and Pharmacological Sciences* 2011; 15(7):775-780.
51. Michael LM, Mazuru G, Nyasha G, Godfred H. Chemical composition and biological activities of essential oil from the leaves of *Sesuvium portulacastrum*. *J Ethnopharmacology* 2006; 103:85-90.
52. Bandaranayake WM. Bioactivities, bioactive compounds and chemical constituents of mangrove plants. *Wetlands Ecol Manage* 2002; 10:421-52.
53. Fauvel MT, Taoubi K, Gleye J, Fouraste I. Phenylpropanoid glycosides from *Avicennia marina*. *Planta Med* 1993; 59(4):387.
54. Gurudeeban S, Satyavani K, Ramanathan T, Balasubramanian T. Antidiabetic effect of a black mangrove species *Aegiceras corniculatum* in alloxan induced diabetic rat. *J Adv Pharm Technol Res* 2012; 3:52-6.
55. Han L, Huang XS, Dahse HM, Moellmann U, Grabley S, Lin WH, *et al.* New abietane diterpenoids from the mangrove *Avicennia marina*. *Planta Med* 2008; 74:432-7.
56. Han L, Huang XS, Dahse HM, Moellmann U, Fu HZ, Grabley S, *et al.* Unusual naphthoquinone derivatives from the twigs of *Avicennia marina*. *J Nat Prod* 2007; 70:923-7.
57. Feng Y, Li XM, Duan XJ, Wang BG. Iridoid glucosides and flavones from the aerial parts of *Avicennia marina*. *Chem Biodivers* 2006; 3:799-806.
58. Feng Y, Li XM, Duan XJ, Wang BG. A new acylated iridoid glucoside from *Avicennia marina*. *Chin Chem Let* 2006; 17:1201-4.
59. Sharma M, Garg HS. Iridoid glycosides from *Avicennia officinalis*. *Ind J Chem* 1996; 35:459-62.
60. A. M. Migahid, *Flora of Saudi Arabia*, Riyadh University, Riyadh, Saudi Arabia, 2nd edition, 1978.

61. A. M. Youssef, M. A. Al-Fredan, and A. A. Fathi, "Floristic composition of lake Al-Asfar, Alahsa, Saudi Arabia," *International Journal of Botany*, vol. 5, no. 2, pp. 116–125, 2009.
62. J. K. Patra, N. K. Dhal, and H. N. Thatoi, "In vitro bioactivity and phytochemical screening of *Suaeda maritima* (Dumort): a mangrove associate from Bhitarkanika, India," *Asian Pacific Journal of Tropical Medicine*, vol. 4, no. 9, pp. 727–734, 2011.
63. W. M. Bandaranayake, "Traditional and medicinal uses of mangroves," *Mangroves and Salt Marshes*, vol. 2, no. 3, pp. 133–148, 1998.
64. S. Singh, S. K. Sharma, and R. Mann, "Pharmacognostical standardization of stem of *Suaeda maritima*(L.) Dumort," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 4, no. 3, pp. 304–306, 2012.
65. S. Ravikumar, M. Gnanadesigan, S. Jacob Inbaneson, and A. Kalaiarasi, "Hepatoprotective and antioxidant properties of *Suaeda maritima* (L.) Dumort ethanolic extract on concanavalin-A induced hepatotoxicity in rats," *Indian Journal of Experimental Biology*, vol. 49, no. 6, pp. 455–460, 2011.
66. S. Singh, R. Mann, and S. K. Sharma, "Pharmacognostical standardization of root of *Suaeda maritima*(L.) dumort," *Der Pharmacia Lettre*, vol. 5, no. 1, pp. 116–120, 2013.
67. Premnathan M, Chandra K, Bajpai SK, Kathiresan K, *et al.* A survey of some Indian marine plants for antiviral activity. *Botanical Marina* 1992; 35: 321-324
68. Ravikumar S, Gnanadesigan M, Inbaneson SJ, Kalaiarasi A, *et al.* Hepatoprotective and antioxidant properties of *Suaeda maritima* (L.) Dumort ethanolic extract on concanavalin-A induced hepatotoxicity in rats. *Indian Journal of Experimental Biology*, 2011; 49: 455-460.
69. Ravikumar S, Gnanadesigan M, Serebiah JS, Inbaneson SJ, *et al.* Hepatoprotective effect of an Indian salt marsh herb *Suaeda monoica* Forrsk ex. Gmel against concanavalin-A induced toxicity in rats. *Life Science Medicinal Research* 2010; 2: 1-9.
70. Kokpal V, Miles DH, Payne AM, Chittarwong V *et al.* Chemical constituents and bioactive compounds from mangrove plants. *Studies in Natural products Chemistry*, 1990; 7: 175-199
71. Bandaranayake WM: Traditional and medicinal uses of mangroves. *Mangroves and Salt Marshes*. 1998, 2(Suppl 3): 133–148.
72. Padmakumar K, Ayyakkannu K: Antiviral activity of marine plants. *Indian Journal of Virology*. 1992, 13: 33–36.
73. Premanathan M, Chandra K, Bajpai SK, Kathiresan K: A survey of some Indian marine plants for antiviral activity. *Botanica Marina* 1992, 35: 321–324.
74. Ghosh A, Misra S, Duta AK, Choudhury A: Pentacyclic triterpinoids and sterols from seven species of mangrove. *Phytochemistry* 1985, 24: 1725-1727.
75. Subramanyam C, Rao KB, Rao CV, Rao BV: Chemical examination of *Suaeda monoica* and *Suaeda maritima*. *Acta Cienca Indica* 1992, 18: 7-8.
76. Ralston L, Suramanian S, Matsuno M, Yu O. Partial reconstruction of flavonoid and isoflavonoid biosynthesis in yeast using soybean type I and type II chalcone isomerases. *Plant Physiol*, 2005; 137:1375-1388.
77. Asha KK, Mathew S, Lakshmanan PT. Flavonoids and phenolic compounds in two mangrove species and their antioxidant property. *Indian J Geo-Marine Sci*, 2012; 41(3): 259-264.
78. Middleton EJ. Effect of plant flavonoids on immune and inflammatory cell function. *Adv*

- Exp Med Biology*, 1998;439:175-182.
79. Cushnie TPT, Lamb AJ. Antimicrobial activity of flavonoids. *Int J Antimicrob Agents*, 2005; 26:343-356.
 80. Ravindran KC, Venkatesan K, Balakrishnan V, Chellappan KP, Balasubramanian. Ethnomedicinal studies of Pichavaram mangroves of East coast, Tamil Nadu. *Indian J Traditional Knowledge*, 2005; 4(4): 409-411.
 81. Manikandan T, Neelakandan T, Rani UG. Antibacterial activity of *Salicornia brachiata*, a halophyte. *J Phytol*, 2009;1(6):441-443.
 82. Stanley OD. Bio prospecting marine halophyte *Salicornia brachiata* for medical importance and salt encrusted land development. *J. Coas. Develop.*, 2008;11(2):62-69.
 83. Kathiresan K, Bingham BL. Biology of mangrove and mangrove ecosystem. *Adv in Marine Biol*, 2001; 40: 81-251.
 84. Rizk AM. The Phytochemistry of the Flora of Qatar. University of Qatar, Doha, Qatar, Scientific and Applied Research Centre. 1986.
 85. Park DI. Methods utilizing pharmacological activities of *Salicornia herbacea*. 2000; Korea Patent 2000-0074066.
 86. Deepa S, Kannan P, Kanth SV, Rao J, Chandrasekaran B. Ramesh Raghava Antioxidant and cytotoxic effects of methanolic extract of *Salicornia brachiata*. *International Journal of Research in Pharmaceutical Sciences* a 2013;4:512-7